Hek293/Hek293-Lif/IMR32/IIC9 cells maintenance

Hek Mixed media:

	50mls	150mls	250mls	500mls
EMEM	44.5	133.5	222.5	445
FBS(final 10%)	5.0	15.0	25.0	50.0
P/S/L-glut	0.5	1.5	2.5	5.0

- Allow the media to warm for about one hour in the 37ºC water bath.
- Cells frozen in DMSO can be thawed by gently shaking by hand in the water bath.
- Gently remove cells from tube with 1mL pipette and transfer to P10.
- Add 10 mLs of media (above)
- Place in incubator
- The following day, remove old media and replace with new to remove remaining DMSO

Notes:
-EMEM (MEM Eagle with Earle's BSS; Description: w/ L-Glutamine): Biowhittaker from Fisher Cat#: BW12-611Q

Split Media (Hek293/Hek293-Lif/IMR32)

-Trypsin should be diluted 4X!

Example: For 5mls use 1.25mls Trypsin with 3.75mls HBSS
-Remove the media and add ½ amount of 4x diluted trypsin as the total amount of media removed. If the cells are on a P10 and 10mls of media was removed add 5mls of trypsin + HBSS.
-Incubate in the incubator for about 1-2 min.
-Blow off cells and place in a 15ml 0r 50ml tube
-Do not spin down
-Dilute by adding media if necessary
-Plate the cells as needed